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(54) Title: AMINO ACID-DERIVED PHOSPHONAMIDIC ANHYDRIDES AND METHODS OF PREPARING THE SAME

(57) Abstract: New phosphonamide compounds and methods of forming those compounds are provided. The inventive methods comprise forming a template opened-ring phosphonamide compound and optionally subjecting the template to a ring-closing metathesis reaction in the presence of a ring-closing catalyst (e.g., a Grubbs catalyst) to yield a heterocyclic phosphonamide. Advantageously, the template structures can be provided with a wide array of functional groups (e.g., amino acid side chains, peptides) chosen to provide particular properties to the compound. The preferred heterocyclic phosphonamides are represented by formula (I).

AMINO ACID-DERIVED PHOSPHONAMIDIC ANHYDRIDES AND METHODS OF PREPARING THE SAME

BACKGROUND OF THE INVENTION

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Field of the Invention

The present invention is directed towards new phosphonamidic anhydrides and methods of forming those compounds. The compounds have a number of uses including as inhibitors of enzymes and regulators of plant growth.

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Description of the Prior Art

Small peptides are excellent starting points for drug design because they have the potential to overcome the pharmacokinetic shortcomings of larger peptides, yet retain the desirable quality of molecular recognition. A number of dipeptides are currently being developed as novel pharmaceutical agents (see e.g., Blackburn et al., *Bioorg. Med. Chem. Lett.*, 7:823-26 (1997); Schullek et al., *Anal. Biochem.*, 246:20-29 (1997), each incorporated by reference herein). Unfortunately, even small peptides suffer from proteolytic instability which limits their use as drug candidates.

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Anhydrides and their derivatives have a rich history both in terms of their synthetic utility as well as their biological relevance (see e.g., Tarbell, *Accounts Chem. Res.*, 2:296-300 (1969); Martin et al., *Chem.*, 27:90-95 (1987), each incorporated by reference herein). Anhydrides are widely known to serve as potent inhibitors of a variety of enzymes (see e.g., Karibian et al., *Biochemistry*, 13:2891 (1974), incorporated by reference herein), with a number of anhydrides recently being reported as effective inactivators of various serine proteases (see e.g., Iijima et al., *Biorg. Med. Chem. Lett.*, 9:413 (1999), incorporated by reference herein).

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Pyrophosphate and related analogs are a class of phosphorus-based anhydrides that have gained attention for their ability to inhibit osteoclastic bone resorption, and therefore are useful therapeutic agents to treat and prevent osteoporosis (see e.g., Sato et al., *J. Med. Chem.*, 42:1 (1999), incorporated by reference herein). Biphosphonates, synthetic nonhydrolyzable P-C-P analogs of pyrophosphates, are highly effective agents for inhibiting osteoclastic bone resorption (see e.g., Russell et al., *Bone*, 25:97 (1999); Teronen et al., *Ann. N. Y. Acad. Sci.*, 878:453-65 (1999), each incorporated by reference herein). Biphosphonic acids have also proven to be effective inhibitors of squalene synthase, a crucial enzyme in the role of cholesterol biosynthesis.

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Thus, there is a need to develop new pyrophosphate analogs which improve the therapeutic properties of biphosphonates.

SUMMARY OF THE INVENTION

The present invention is broadly concerned with new phosphonamide compounds and methods of forming such compounds.

In more detail, the compounds are phosphonamidic anhydrides, and more particularly chiral phosphonamidic anhydrides. The preferred compounds are represented by a formula selected from the group consisting of

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$$R^{2}XC \xrightarrow{R^{1}} \begin{array}{c} R^{2} & Q & R^{2} & Q & R^{1} \\ & & & & & & & & \\ & & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & & \\ & & & & & \\ & & & & \\ & & & & & \\ & & & & \\ & & & & & \\ & & & & & \\ & & & \\ & & & & \\ & & &$$

and

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$$P$$
 P
 O
 P
 N
 P
 O
 P
 N
 R^2O_2C
 R^1
 R^1
 R^1
 CO_2R^2

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wherein:

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each X is individually selected from the group consisting of oxygen, -NH, and -NOR¹; each R¹ is individually selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

each R^2 is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups (preferably C_1 - C_{18} , more preferably C_1 - C_8), branched and unbranched alkenyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_{18} , more preferably C_2 - C_{18}), branched and unbranched alkynyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_{18}).

 C_8), allyl groups, acyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8) aryl groups (preferably C_6 - C_{12}), 2-15 mer peptides, and benzyl groups.

Preferably at least one R¹ group comprises an amino acid side chain selected from the group consisting of

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wherein each R^3 is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups (preferably C_1 - C_{18} , more preferably C_1 - C_8), branched and unbranched alkenyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), branched and unbranched alkynyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), allyl groups, aryl groups (preferably C_6 - C_{12}), acyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), and benzyl groups.

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In a preferred embodiment, each R^1 is individually selected from the group consisting of -CH₃, -CH₂CH(R^4)₂, -CH₂ R^4 , and -CH(R^4)₂, with each R^4 being individually selected from the

group consisting of alkyl groups (preferably methyl), aryl groups (preferably phenyl), and benzyl groups, and each R^2 is individually selected from the group consisting of $-CH_3$ and $-CHCH_2$. Two particularly preferred compounds according to the invention comprise a formula selected from the group consisting of

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The inventive compounds are formed by reacting an allylated compound with a phosphonic compound to form an intermediate compound which is the dimerized. Preferred allylated compounds comprise the formula

 R^1 OR^2

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wherein:

R¹ is selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

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 R^2 is selected from the group consisting of hydrogen, branched and unbranched alkyl groups (preferably C_1 - C_{18} , more preferably C_1 - C_8), branched and unbranched alkenyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), branched and unbranched alkynyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), allyl groups, aryl groups (preferably C_6 - C_{12}), acyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), 2-15 mer peptides, and benzyl groups.

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Preferred phosphonic compounds comprise the formula R⁴POY₂,

wherein:

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R⁴ is selected from the group consisting of hydrogen, branched and unbranched alkyl groups (preferably C₁-C₁₈, more preferably C₁-C₈), branched and unbranched alkenyl groups (preferably C₂-C₁₈, more preferably C₂-C₈), branched and unbranched alkynyl groups (preferably C₂-C₁₈, more preferably C₂-C₈), allyl groups, aryl groups (preferably C₆-C₁₂), acyl groups (preferably C₂-C₁₈, more preferably C₂-C₈), 2-15 mer peptides, and benzyl groups; and

each Y is individually selected from the group consisting of the halogens.

The intermediate compound comprises a formula selected from the group consisting of

and

wherein:

R¹ is selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides;

 R^2 is selected from the group consisting of hydrogen, branched and unbranched alkyl groups (preferably C_1 - C_{18} , more preferably C_1 - C_8), branched and unbranched alkenyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), branched and unbranched alkynyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), allyl

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groups, aryl groups (preferably C_6 - C_{12}), acyl groups (preferably C_2 - C_{18} , more preferably C_2 - C_8), 2-15 mer peptides, and benzyl groups; and

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each Y is individually selected from the group consisting of the halogens.

Preparing the phosphonamide compounds according to the inventive methods results in a yield of those compounds of at least about 70%, and preferably at least about 95%, wherein the theoretical yield is taken as 100%.

Optionally, the phosphonamide compound can be subjected to a ring-closing metathesis reaction in the presence of a ring-closing catalyst to yield a bicyclic phosphonamide. Preferred ring-closing catalysts are olefin metathesis catalysts such as Grubbs catalysts (see e.g., U.S. Patent Nos. 6,048,993, 5,917,071, 5,750,815, 5,710,298, 5,342,909, and 5,312,940, each incorporated by reference herein) as well as those disclosed by the following references, each also incorporated by reference herein: Matthias, *Org. Ltrs.*, 1(6):953-56 (1999); Schrock, *Macromolecules*, 29(19):6114-25 (1996); Zhu et al., *J. Amer. Chem. Soc.*, 121(36):8251-59 (1999); Alexander et al., *J. Amer. Chem. Soc.*, 120(16):4041-42 (1998); and Kingsbury et al., *J. Amer. Chem. Soc.*, 121(4):791-99 (1999).

Particularly preferred Grubbs catalysts are those selected from the group consisting of

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$$\begin{array}{c|c}
CI & PCy_3 \\
Ru & Ph \\
PCy_3 & CI & Ru \\
PCy_3 & CI & PCy_3
\end{array}$$
and
$$\begin{array}{c|c}
CI & PCy_3 \\
Ph \\
CI & PCy_3
\end{array}$$
and

Preferably the reacting step is carried out at a temperature of from about 15-80°C, and more preferably from about 30-55°C. Furthermore, the reacting step should be carried out in a solvent system comprising a solvent selected from the group consisting of toluene, benzene,

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chlorobenzene, dichlorobenzene, methylene chloride, dimethoxyethane (DME), and mixtures thereof.

It will be appreciated that the inventive methods allow for the synthesis of a wide array of both symmetric and unsymmetric cyclic and acyclic phosphonamide compounds. Furthermore, the inventive methods allow for preparation of, or selection of, templates having particular functional groups bonded thereto which are then readily formed into the desired phosphonamide in a controlled and repeatable manner. Because the method can be adapted to form phosphonamide compounds comprising one or more amino acid side chains or peptides bonded thereto, the inventive compounds can be used to inhibit enzymes (such as squalene synthetase), to act as osteoporitic agents, and to regulate plant growth.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS EXAMPLES

The following examples set forth preferred methods in accordance with the invention. It is to be understood, however, that these examples are provided by way of illustration and nothing therein should be taken as a limitation upon the overall scope of the invention.

A number of abbreviations are used herein. These abbreviations and the term or terms that they represent are set forth in Table A.

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Table A

Abbreviation	Term(s)	
hex	hexane	
Bn	benzyl	
Ph	phenyl	
Me	methyl	
Et	ethyl	
Вос	butoxy carbonyl	
EtOAc	ethyl acetate	
Et ₃ N	triethyl amine	
^t BuO	tert-butoxy	

Grubbs Catalysts were used in some of the following Examples. These catalysts are referred to as follows:

$$\begin{array}{c|c}
Cl & PCy_3 \\
Cl & Ru \\
PCy_3
\end{array}$$
Ph
$$\begin{array}{c|c}
PCy_3 \\
Cl & Ru \\
PCy_3
\end{array}$$
Ph
$$\begin{array}{c|c}
PCy_3 \\
PCy_3
\end{array}$$
Ph

Grubbs Catalyst 1

Grubbs Catatlyst 2

Grubbs Catalyst 3

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EXAMPLE 1

Scheme A depicts the general overall reaction scheme followed in Parts I-II below, as well as the various compounds which can be prepared according to the procedure described in this example.

Scheme A

OME DMAP, Et₃N
$$CH_2Cl_2$$
 R^2POCl_2 $23: R^2 = CH_3$ $24: R^2 = CHCH_2$ $25: R^1 = CH_2CHMe_2; R^2 = CH_3$ $26: R^1 = CH_2CHMe_2; R^2 = CH_3$ $27: R^1 = CH_2Ph; R^2 = CH_3$ $28: R^1 = CH_3Ph; R^2 = CH_3$ $29: R^1 = CH_3; R^2 = CHCH_2$ $29: R^1 = CH_2Ph; R^2 = CHCH_2$ $20: R^1 = CH_2Ph$ $20: R^1 = CH_2Ph$ $20: R^1 = CH_2Ph$ $20: R^1 = CH_2Ph; R^2 = CHCH_2$ $20: R^1 = CH_2Ph; R^2 = CH_2Ph; R^2$

 $25-32-P_RS$

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In this and the following procedure descriptions, the number/letter abbreviation depicted in the particular reaction scheme follows the chemical name of the particular compound (e.g., "(20)" follows "allylated leucine methyl ester").

I. Preparation of Leucine-Derived Methyl Phosphonamidic Chloridates ($26P_sS$ and $26P_sS$)

Methylphosphonic dichloride (23) (1.0 mL, 11.04 mmol) and CH_2Cl_2 (20 mL) were added to a flame-dried 100 mL round bottom flask under argon atmosphere. The reaction flask was cooled to 0°C, and Et_3N (6.26 mL, 45.0 mmol) was added dropwise, followed by a catalytic amount of 4-dimethylaminopyridine (DMAP) (5 mol %). After stirring at 0°C for 5 minutes, 0.98 equivalents of allylated leucine methyl ester (20) (2.0 g, 10.82 mmol) in CH_2Cl_2 (5 mL) was added via cannulae. The reaction mixture was refluxed and monitored by TLC. Once complete, the reaction mixture was concentrated under reduced pressure, diluted with EtOAc, filtered, and further concentrated under reduced pressure. Flash chromatography (SiO_2 , 3:1 Hex/EtOAc) gave 2.94g (95%) of a light yellow oil consisting solely of the two diastereomeric chloridates ($26P_sS$) and ($26P_RS$) (see Scheme B). Further chromatography (SiO_2 , 8:1 Hex/EtOAc) yielded portions of the separated isomers for characterization.

Scheme B

The leucine-derived methyl phosphonamidic chloridate ($26P_sS$ or $26P_RS$, top R_f) was characterized as follows: TLC $R_f = 0.39$ (1:1 Hex:EtOAc); $[\alpha]^{25} = -42.3$ (c = 2.44, CHCl₃);. FTIR 1742, 1445, 1368, 1240 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.78 (dddd, 16.9, 10.2, 6.5, 6.5 Hz, 1H), 5.22 (dd, J = 17.2, 1.2 Hz, 1H), 5.14 (d, J = 10.1 Hz, 1H), 4.40 (ddd, $J_{HP} = 12.1$ Hz, $J_{HH} = 7.5$, 7.5 Hz, 1H), 3.75-3.67 (m, 2H), 3.68 (s, 3H), 1.96 (d, $J_{HP} = 16.3$ Hz, 3H), 1.70 (dd,

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J= 7.3, 6.3 Hz, 2H), 1.65-1.52 (m, 1H), 0.91 (d, J= 6.4 Hz, 3H), 0.90 (d, J= 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.37, 134.68 (d, $J_{\rm CP}$ = 3.0 Hz), 118.04, 55.87, 52.05, 47.16 (d, $J_{\rm CP}$ = 4.5 Hz), 38.49 (d, $J_{\rm CP}$ = 5.7 Hz), 24.52, 22.65 (d, $J_{\rm CP}$ = 118.9 Hz), 22.65, 21.53; ³¹P NMR (162 MHz, CDCl₃) δ 48.02; HRMS calculated for C₁₁H₂₃ClNO₃P (M+H)⁺ required 282.1026, found 282.1049.

The leucine-derived methyl phosphonamidic chloridates ($26P_RS$ or $26P_SS$, bottom R_f) were characterized as follows: TLC R_f = 0.38 (1:1 Hex:EtOAc); [α]²⁵ = -13.1 (c = 1.44, CHCl₃); FTIR 1742, 1440, 1373, 1245 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.81-5.69 (m, 1H), 5.20-5.08 (m, 2H), 4.51 (ddd, J_{HP} = 9.2 Hz, J_{HH} = 6.2, 6.2 Hz, 1H), 3.76-3.66 (m, 2H), 3.62 (s, 3H), 1.98 (d, J_{HP} = 16.0 Hz, 3H), 1.73-1.64 (m, 2H), 1.64-1.51 (m, 1H), 0.89 (d, J_{CP} = 6.3 Hz, 3H), 0.88 (d, J_{CP} = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.56 (d, J_{CP} = 6.2 Hz), 134.08 (d, J_{CP} = 2.8 Hz), 118.06, 55.35 (d, J_{CP} = 2.0 Hz), 51.97, 46.47 (d, J_{CP} = 5.1 Hz), 37.14 (d, J_{CP} = 2.1 Hz), 24.31, 22.80, 22.28 (d, J_{CP} = 117.3 Hz), 21.20; ³¹P NMR (162 MHz, CDCl₃) δ 47.98; HRMS calculated for $C_{11}H_{23}$ ClNO₃P (M+H)⁺ required 282.1026, found 282.1047.

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II. Preparation of Leucine-Derived Vinyl Phosphonamidic Chloridates (30P_sS and 30P_pS) CH₂Cl₂ (36 mL) and vinylphosphonic acid (1 mL, 12.86 mmol) were added to a flamedried 100 mL round bottom flask under an inert atmosphere. The reaction mixture was stirred vigorously, and oxalyl chloride (3.36 mL, 38.6 mmol) was added followed by the addition of a catalytic amount of dimethyl formamide (DMF) (1 drop). The system was stirred for 2 hours (until gas evolution was no longer apparent). Upon completion of the reaction, the mixture was concentrated under reduced pressure to yield the vinylphosphonic dichloride (24) as a yellow oil. This oil was diluted with CH₂Cl₂ (20 mL) while precaution was taken to avoid exposure to moisture. The mixture was then cooled to 0°C and Et₃N (6.26 mL, 45.0 mmol) was added dropwise followed by DMAP (5 mol %). After stirring the reaction mixture at 0°C for 5 minutes, allylated leucine methyl ester (20) (2.38g, 12.86 mmol) in CH₂Cl₂ (5 mL) was added via cannulae. The reaction mix was refluxed and monitored by TLC. Once complete, the reaction mixture was concentrated under reduced pressure, diluted with EtOAc, filtered, and further concentrated under reduced pressure. Flash chromatography (3:1 Hexane/EtOAc) gave the two diastereomeric leucine-derived phosphonamidic chloridates ($30P_sS$ and $30P_pS$ (3.58 g,

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95%) (see Scheme C) as a light yellow oil. These phosphonamidic chloridates were not further characterized.

Scheme C

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In Parts I-II of this procedure, methylene chloride was the solvent utilized. However, acetonitrile, chloroform, toluene, benzene, tetrahydrofuran (THF), diethyl ether, dimethoxyethane (DME), and mixtures thereof are also suitable solvents. Furthermore, while Et₃N was used as the base, other bases which could be used include pyridine, NaHCO₃, Na₂CO₃, K₂CO₃, NaH, KH, any tertiary amine, and mixtures thereof. Finally, while the procedure was carried out at temperatures of 0-20 °C, temperatures of anywhere from about -20-20 °C would also be suitable.

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EXAMPLE 2

Scheme D depicts the general overall reaction scheme followed in Parts I-VIII below, as well as the various compounds which can be prepared according to the procedure described in this example.

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Scheme D

$$MeO_2C$$
 N
 P
 O
 P
 N
 CO_2Me
 SP_RP_SS

Pseudo-Meso

4-11c

I. Preparation of Alanine-Derived Methyl Phosphonamidic Anhydrides (4a-c)

A neat mixture of the diastereomeric leucine phosphonamidic chloridates (25 P_RS) and (25 P_SS) (500 mg, 2.09 mmol) was allowed to sit at ambient temperature over a period of 2-4 days. The slurry was partitioned between EtOAc and water. The water layer was re-extracted with EtOAc, and the organic layers were combined, dried with Na₂SO₄, and concentrated under reduced pressure to leave a crude oil. Flash chromatography (SiO₂, EtOAc) afforded 120 mg (27%) of the pseudo-*meso* diastereomer (4c) and 137 mg (31%) of an inseparable mixture of C_2 -symmetric diastereomers (4a) and (4b), both as colorless oils. Scheme E depicts each of these compounds.

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Scheme E

$$CH_3 \text{ Me } O \text{ Me } O \text{ CH}_3$$

$$SP_SP_SS$$

$$C_2\text{-Symmetric}$$

$$Aa$$

$$CH_3 \text{ Me } O \text{ Me } O \text{ CH}_3$$

$$CO_2Me$$

$$CO_2Me$$

$$CH_3 \text{ Me } O \text{ Me } O \text{ CH}_3$$

$$C_2\text{-Symmetric}$$

$$CH_3 \text{ Me } O \text{ Me } O \text{ CH}_3$$

$$C_2\text{-Symmetric}$$

$$Ab$$

$$CO_2Me$$

$$SP_RP_SS$$

$$C_2\text{-Symmetric}$$

$$Ab$$

$$CO_2Me$$

$$CO_2Me$$

$$CO_2Me$$

$$CO_2Me$$

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The C_2 -symmetric alanine-derived methyl phosphonamidate anhydrides (**4a,b**) were characterized, as a mixture, as follows: TLC R_f = 0.10 (EtOAc); FTIR 2992, 2951, 1740, 1457, 1437, 1382, 1232, 1170 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.82-5.72 (m, 4H, mix), 5.23-5.07 (m, 8H, mix), 4.54-4.48 (m, 2H, mix), 4.42-4.33 (m 2H, mix), 3.79-3.61 (m, 8H, mix), 3.67 (s,

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6H), 3.66 (s, 6H), 1.73 (d, J_{HP} = 16.6 Hz, 6H), 1.71 (d, J_{HP} = 16.9 Hz, 6H), 1.43 (d, J = 7.2 Hz, 6H), 1.42 (d, J = 7.3 Hz, 6H); 13 C NMR (100 MHz, CDCl₃) δ 173.46, 173.20, 135.71, 135.33, 117.30, 117.08, 53.49, 52.99, 52.02, 52.02, 46.52, 46.32, 16.97, 16.10, 15.53 (dd, J_{CP} = 137.7, 6.2 Hz), 14.99 (dd, J_{CP} = 136.6, 4.6 Hz); 31 P NMR (162 MHz, CDCl₃) δ 29.47, 29.41; HRMS calculated for $C_{16}H_{31}N_2O_7P_2$ (M+H)⁺ required 425.1614, found 425.1612.

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The pseudo-*meso* alanine-derived methyl phosphonamidic anhydride (**4c**) was characterized as follow: TLC R_f = 0.15 (EtOAc); $[\alpha]^{25}$ = -0.33 (c = 0.60, CHCl₃); FTIR 2989, 2950, 1740, 1457, 1437, 1381, 1241 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.84- 5.72 (m, 2H), 5.18 (dd, J = 17.3, 1.3 Hz, 1H), 5.16 (dd, J = 17.2, 1.3 Hz, 1H), 5.10-5.05 (m, 2H), 4.52 (dq, J_{HP} = 11.1 Hz, J_{HH} = 7.3 Hz, 1H), 4.23 (dq, J_{HP} = 14.3 Hz, J_{HH} = 7.1 Hz, 1H), 3.75-3.59 (m, 4H), 3.66 (s, 3H), 3.65 (s, 3H), 1.72 (d, J_{HP} = 16.5 Hz, 3H), 1.66 (d, J_{HP} = 16.6 Hz, 3H), 1.45 (d, J = 7.2 Hz, 3H), 1.41 (d, J = 7.3 Hz, 3H); 13 C NMR (100 MHz, CDCl₃) δ 173.48, 173.08, 135.35, 135.35, 117.40, 117.32, 53.90, 53.06, 52.02, 51.98, 47.14, 46.25, 17.05, 15.96, 15.23 (dd, J_{CP} = 140.2, 6.5 Hz), 14.77 (dd, J_{CP} = 139.6, 6.9 Hz); 31 P NMR (162 MHz, CDCl₃) δ 28.95 (d, J_{PP} = 34.4 Hz), 28.94 (d, J_{PP} = 34.4 Hz); HRMS calculated for C₁₆H₃₁N₂O₇P₂ (M+H)⁺ required 425.1614, found 425.1606.

II. Preparation of Leucine-Derived Methyl Phosphonamidic Anhydrides (5a-c)

Et₃N (450 μ L, 3.22 mmol) was added to a neat solution of a mixture of the diastereomeric leucine phosphonamidic chloridates (26P_RS) and (26P_SS) (260 mg, 0.92 mmol) at 0°C. The mixture was heated at 45°C and monitored by TLC and ³¹P NMR. The resulting salty slurry was diluted with EtOAc (10 mL), filtered (10 mL), and concentrated under reduced pressure to yield 236 mg (quantitative) of a mixture of the three diastereomeric anhydrides as a yellow oil. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 46 mg (20%) of the pseudo-*meso* diastereomer (5c) and 132 mg (56%) of a mixture of the C_2 -symmetric diastereomers (5a) and (5b). The mixture comprised of 16 mg (7%) of a single C_2 -symmetric diastereomer (5a) or (5b), 104 mg (44%) of a mixture of C_2 -symmetric diastereomers (5a) and (5b), and 12 mg (5%) of a sample of C_2 -symmetric diastereomer (5b) or (5a) at 90% purity, all as colorless oils.

The pseudo-*meso* leucine-derived methyl phosphonamidic anhydride (**5c**) was characterized as follows: TLC $R_f = 0.4$ (EtOAc); $[\alpha]^{25} = -17.06$ (c = 0.59, CHCl₃); FTIR 1749, 1714, 1422, 1363, 1222 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.87-5.75 (m, 2H), 5.18 (dd,

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J=17.1, 1.3 Hz, 1H), 5.14 (dd, J=17.1, 1.3 Hz, 1H), 5.12-5.06 (m, 2H), 4.49 (ddd, $J_{\rm HP}=10.7$ Hz, $J_{\rm HH}=10.7, 5.4$ Hz, 1H), 4.34 (ddd, $J_{\rm HP}=9.2$ Hz, $J_{\rm HH}=9.2, 5.3$ Hz, 1H), 3.75-3.58 (m, 4H), 3.68 (s, 3H), 3.66 (s, 3H), 1.83-1.60 (m, 6H), 1.74 (d, $J_{\rm HP}=16.8$ Hz, 3H), 1.69 (d, $J_{\rm HP}=16.7$ Hz, 3H), 0.92 (d, J=6.2 Hz, 6H), 0.91 (d, J=6.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 173.67 (d, $J_{\rm CP}=2.2$ Hz), 173.20, 135.34, 135.34, 117.77, 117.48, 56.32 (d, $J_{\rm CP}=2.9$ Hz), 55.93 (d, $J_{\rm CP}=3.6$ Hz), 51.94, 51.90, 47.24 (d, $J_{\rm CP}=4.9$ Hz), 46.45 (d, $J_{\rm CP}=4.9$ Hz), 38.98 (d, $J_{\rm CP}=3.7$ Hz), 37.57 (d, $J_{\rm CP}=2.6$ Hz), 24.51, 24.19, 22.95, 22.87, 21.51, 21.33, 15.49 (dd, $J_{\rm CP}=129.5, 4.3$ Hz), 14.80 (dd, $J_{\rm CP}=127.7, 4.5$ Hz); ³¹P NMR (162 MHz, CDCl₃) δ 29.19 (d, $J_{\rm PP}=35.5$ Hz), 28.41(d, $J_{\rm PP}=35.5$ Hz); HRMS calculated for $C_{23}H_{43}N_2O_7P_2$ (M+H)⁺ required 509.2546, found 509.2545.

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The C_2 -symmetric leucine-derived methyl phosphonamidic anhydride, single diastereomer (**5a** or **5b**, top R_f) was characterized as follows: TLC R_f = 0.22 (EtOAc); $[\alpha]^{25}$ = -37.8 (c = 0.32, CHCl₃); FTIR 1740, 1437, 1387, 1241 (P=O) cm⁻¹; 1 H NMR (400 MHz, CDCl₃) δ 5.79 (dddd, J= 16.9, 10.2, 6.2, 6.2 Hz, 2H), 5.21 (dd, J= 15.9, 1.2 Hz, 2H), 5.12 (dd, J= 9.8, 0.9 Hz, 2H), 4.49-4.42 (m, 2H), 3.79-3.57 (m, 4H), 3.67 (s, 6H), 1.81-1.59 (m, 6H), 1.75 (d, $J_{\rm HP}$ = 17.0 Hz, 6H), 0.94 (d, J= 6.0 Hz, 6H), 0.93 (d, J= 6.2 Hz, 6H); 13 C NMR (100 MHz, CDCl₃) δ 173.33, 135.59, 117.62, 56.11, 51.95, 46.95, 38.71, 24.57, 22.94, 21.31, 15.67 (dd, $J_{\rm CP}$ = 130.9, 5.6 Hz); 31 P NMR (162 MHz, CDCl₃) δ 29.88; HRMS calculated for $C_{23}H_{43}N_2O_7P_2$ (M+H)⁺ required 509.2546, found 509.2526.

The C_2 -symmetric leucine-derived methyl phosphonamidic anhydride, single diastereomer (**5b** or **5a**, bottom R_f at 90% purity) was characterized as follows: TLC R_f = 0.22 (EtOAc); [α]²⁵ = -5.0 (c = 0.24, CHCl₃); FTIR 1740, 1437, 1387, 1241 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.79 (dddd, J= 16.8, 10.1, 6.6, 6.6 Hz, 2H), 5.17 (dd, J= 16.7, 1.3 Hz, 2H), 5.10 (dd, J= 9.1, 1.0 Hz, 2H), 4.52-4.42 (m, 2H), 3.66-3.54 (m, 4H), 3.68 (s, 6H), 1.80 (d, J_{HP} = 15.8 Hz, 6H), 1.78-1.53 (m, 6H), 0.94 (d, J= 6.0 Hz, 6H), 0.93 (d, J= 6.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 173.48, 135.13, 117.47, 55.65, 51.91, 46.35, 37.46, 24.34, 22.99, 21.21, 15.27 (dd, J_{CP} = 130.9, 5.6 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 30.14 HRMS calculated for $C_{23}H_{43}N_2O_7P_2$ (M+H)⁺ required 509.2546, found 509.2561.

In this part of this example, Et₃N was used as the base. However, other bases could be used, including pyridine, NaHCO₃, Na₂CO₃, K₂CO₃, NaH, KH, any tertiary amine, and mixtures thereof. Finally, while the procedure was carried out at temperatures of 0-20°C, temperatures of from about -20-20°C would also be suitable.

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III. Preparation of Phenylalanine-Derived Methyl Phosphonamidic Anhydrides (6a-c)

A mixture of the diastereomeric vinyl chloridates $(27P_SS)$ and $(27P_RS)$ (820 mg, 2.60 mmol) was subjected to the conditions described in Part I of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 195 mg (26%) of the pseudo-*meso* diastereomer (6c) and 172 mg (23%) of an inseparable mixture of C_2 -symmetric diastereomers (6a) and (6b), both as colorless oils.

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The pseudo-*meso* phenylalanine-derived methyl phosphonamidic anhydride (**6c**) was characterized as follows: TLC R_f = 0.4 (EtOAc); $[\alpha]^{25}$ = -11.79 (c = 0.60, CHCl₃); FTIR 1739, 1455, 1437, 1379, 1241, 1169 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.15 (m, 10H), 5.80-5.65 (m, 2H), 5.19 (dd, J = 17.1, 0.9 Hz, 2H), 5.11 (d, J = 10.1 Hz, 2H), 4.68 (ddd, J_{HP} = 12.9 Hz, J_{HH} = 9.6, 5.9 Hz, 1H), 4.64 (ddd, J_{HP} = 13.4 Hz, J_{HH} = 10.0, 6.0 Hz, 1H), 3.69 (s, 3H), 3.68 (s, 3H), 3.65-3.61 (m, 2H), 3.61-3.42 (m, 2H), 3.35 (ddd, J_{HH} = 14.3, 5.7 Hz, J_{HP} = 5.7 Hz, 2H), 3.09 (ddd, J = 14.4, 9.9, 2.8 Hz, 2H), 1.59 (d, J_{HP} = 16.9 Hz, 3H), 0.97 (d, J = J_{HP} = 16.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.41, 172.25, 137.62, 137.51, 134.75, 134.59, 129.35, 129.31, 128.38, 128.36, 128.26, 126.57, 126.50, 118.25, 117.98, 59.03 (d, J_{CP} = 4.0 Hz), 58.96 (d, J_{CP} = 4.1 Hz), 52.03, 51.96, 47.05 (d, J_{CP} = 5.4 Hz), 46.90 (d, J_{CP} = 4.9 Hz), 35.94, 35.30, 14.92 (d, J_{CP} = 128.0 Hz), 14.00 (d, J_{CP} = 126.6 Hz); ³¹P NMR (162 MHz, CDCl₃) 28.96 (d, J_{PP} = 36.7 Hz), 28.12 (d, J_{PP} = 36.7 Hz); HRMS calculated for C₂₈H₃₉N₂O₇P₂ (M+H)⁺ required 577.2233, found 577.2232.

The C_2 -symmetric phenylalanine-derived methyl phosphonamidic anhydride, single diastereomer (**6a** or **6b**, top R_f) was characterized as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.15 (EtOAc); FTIR 1746, 1456, 1437, 1313, 1218, 737, 702 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.18 (m, 10H), 5.75-.35 (m, 2H), 5.17-5.04 (m, 2H), 4.65-4.54 (m, 2H), 3.71-3.57 (m, 4H), 3.69 (s, 6H), 3.33-3.26 (m, 2H), 3.14-3.08 (m, 2H), 1.53 (d, J_{HP} = 17.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 172.41, 137.45, 134.28, 129.12, 128.20, 126.27, 117.90, 58.33, 51.87, 46.90, 34.92, 15.13 (dd, J_{CP} = 135.7, 4.3 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 29.36; LRMS calculated for C₂₈H₃₉N₂O₇P₂ (M+H)⁺ required 577.6, found 577.6.

The C_2 -symmetric phenylalanine-derived methyl phosphonamidate anhydride, single diastereomer (**6a** or **6b**, bottom R_f) was characterized as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC $R_f = 0.15$ (EtOAc); FTIR

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1746, 1456, 1437, 1313, 1218, 737, 702 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.35-7.18 (m, 10H), 5.75-5.35 (m, 2H), 5.17 (d, J= 17.1 Hz, 2H), 5.10 (d, J= 10.5 Hz, 2H), 4.68-4.61 (m, 2H), 3.71-3.57 (m, 4H), 3.66 (s, 6H), 3.38-3.33 (m, 2H), 3.15-3.11 (m, 2H), 1.48 (d, J_{HP} = 17.0 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 172.23, 137.45, 134.48, 129.08, 128.12, 126.44, 118.08, 58.90, 51.92, 47.40, 36.12, 14.96 (dd, J_{CP} = 134.6, 4.4 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 30.10; LRMS calculated for $C_{28}H_{39}N_2O_7P_2$ (M+H)⁺ required 577.6, found 577.6.

IV. Preparation of Valine-derived Methyl Phosphonamidic Anhydrides (7a-c)

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A mixture of the diastereomeric vinyl chloridates ($28P_sS$) and ($28P_RS$) (505 mg, 1.89 mmol) was subjected to the conditions described in Part I of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 91 mg (20%) of the pseudo-*meso* diastereomer (7c) and 195 mg (43%) of an inseparable mixture of C_2 -symmetric diastereomers (7a) and (7b), both as colorless oils.

The characterization of the pseudo-*meso* valine-derived methyl phosphonamidic anhydride (7c) was as follows: TLC $R_f = 0.20$ (EtOAc); $[\alpha]^{25} = -60.51$ (c = 0.43, CHCl₃); FTIR 1739, 1437, 1371, 1311, 1246, 1204 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.86-5.71 (m, 2H), 5.14 (dd, J = 17.1, 1.3 Hz, 2H), 5.09-5.05 (m, 2H), 3.96-3.85 (m, 2H), 3.81-3.71 (m, 2H), 3.69-3.61 (m, 2H), 3.67 (s, 3H), 3.66 (s, 3H), 2.33-2.16 (m, 2H), 1.67 (d, $J_{HP} = 16.5$ Hz, 3H), 1.65 (d, $J_{HP} = 16.6$ Hz, 3H), 0.97 (d, J = 6.7 Hz, 3H), 0.93 (d, J = 6.7 Hz, 3H), 0.89 (d, J = 6.6 Hz, 3H), 0.88 (d, J = 6.5 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.12, 171.79, 135.18, 135.08, 117.67, 117.45, 63.51, 63.49, 51.57, 51.49, 46.44 (d, $J_{CP} = 3.9$ Hz), 45.92 (d, $J_{CP} = 3.3$ Hz), 27.59 (d, $J_{CP} = 3.0$ Hz), 26.81, 19.72, 19.62, 19.39, 19.39, 15.60 (dd, $J_{CP} = 132.7$, 8.8 Hz), 15.63 (dd, $J_{CP} = 134.8$, 8.4 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 29.19 (d, $J_{PP} = 35.4$ Hz), 28.4119 (d, $J_{PP} = 35.4$ Hz). HRMS calculated for $C_{20}H_{30}N_2O_2P_2$ (M+H)⁺ required 481.2233, found 481.2234.

Characterization of the C_2 -symmetric valine-derived methyl phosphonamidate anhydride, single diastereomer (7a or 7b, top R_f) was as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.10 (EtOAc); FTIR 1739, 1436, 1370, 1309, 1245, 1204 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.76-5.61 (m, 2H), 5.10-5.01 (m, 4H), 3.86-3.80 (m, 2H), 3.78-3.42 (m, 4H), 3.60 (s, 6H), 2.25-2.13 (m, 2H), 1.70 (d, J_{HP} = 16.7 Hz, 3H), 1.67 (d, J_{HP} = 16.8 Hz, 3H), 0.90-0.83 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 172.17, 134.91, 117.67, 63.37, 51.65, 45.91, 27.75, 26.91, 19.79, 16.03 (dd, J_{CP} = 133.6, 3.8 Hz); ³¹P

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NMR (162 MHz, CDCl₃) δ 30.20; LRMS calculated for $C_{20}H_{39}N_2O_7P_2$ (M+H)⁺ required 480.1, found 480.1.

Characterization of the C_2 -symmetric valine-derived methyl phosphonamidate anhydride, single diastereomer (7a or 7b, bottom R_f) was as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC $R_f = 0.10$ (EtOAc); FTIR 1739, 1436, 1370, 1309, 1245, 1204 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.84-5.73 (m, 2H), 5.19 (dd, J = 15.9, 1.1 Hz, 2H), 5.12 (d, J = 8.8 Hz, 2H), 3.95-3.89 (m, 2H), 3.83-3.74 (m, 2H), 3.71-3.58 (m, 2H), 3.68 (s, 6H), 2.33-2.20 (m, 2H), 1.78 (d, $J_{HP} = 16.9$ Hz), 1.75 (d, $J_{HP} = 17.1$ Hz), 0.98-0.91 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 172.16, 135.25, 117.77, 63.24, 51.62, 46.37, 27.75, 26.91, 19.72, 15.32 (dd, $J_{CP} = 134.3$, 4.8 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 29.94; LRMS calculated for $C_{20}H_{39}N_2O_7P_2$ (M+H)⁺ required 480.1, found 480.1.

V. Preparation of Alanine-Derived Vinyl Phosphonamidic Anhydrides (8a-c)

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A mixture of the diastereomeric vinyl chloridates $(29P_SS)$ and $(29P_RS)$ (1.22g, 4.86 mmol) was subjected to the conditions described in Part I of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 338 mg (31%) of the pseudo-*meso* diastereomer (8c) and 218 mg (20%) of an inseparable mixture of C_2 -symmetric diastereomers (8a) and (8b), both as colorless oils.

The characterization of the pseudo-*meso* alanine-derived vinyl phosphonamidic anhydride (8c) was as follows: TLC R_f = 0.20 (EtOAc); $[\alpha]^{25}$ = 2.93 (c = 0.82, CHCl₃); FTIR 1738, 1448, 1382, 1221, 1170 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.38-6.22 (m, 4H), 6.19-6.12 (m, 1H), 6.10-5.99 (m 1H), 5.82-5.69 (m, 2H), 5.14 (dd, J = 17.1, 1.3 Hz, 1H), 5.11 (dd, J = 17.2, 1.3 Hz, 1H), 5.07-5.02 (m, 2H), 4.45 (dq, J_{HP} = 12.0 Hz, J_{HH} = 7.3 Hz, 1H), 4.20 (dq, J_{HP} = 14.4 Hz, J_{HH} = 7.2 Hz, 1H), 3.70-3.62 (m, 4H), 3.64 (s, 3H), 3.63 (s, 3H), 1.44 (d, J = 7.3 Hz, 3H), 1.40 (d, J = 7.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 173.31 (d, J_{CP} = 2.8 Hz), 173.00, 135.25, 135.10, 133.96, 133.64, 128.21 (dd, J_{CP} = 180.3, 5.8 Hz), 127.09 (dd, J_{CP} = 181.3, 5.2 Hz), 117.62, 117.58, 53.69 (d, J_{CP} = 3.7 Hz), 52.79 (d, J_{CP} = 4.1 Hz), 51.93, 51.89, 47.17 (d, J_{CP} = 4.3 Hz), 46.41 (d, J_{CP} = 4.3 Hz), 16.90, 15.76; ³¹P NMR (162 MHz, CDCl₃) δ 16.40 (d, J_{PP} = 34.5 Hz), 16.25 (d, J_{PP} = 34.5 Hz); HRMS calculated for C₁₈H₃₁N₂O₇P₂ (M+H)⁺ required 449.1607, found 449.1613.

The characterization of the C_2 -symmetric alanine-derived vinyl phosphonamidic anhydride, single diastereomer (8a or 8b, top R_f) was as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.10 (EtOAc); FTIR 1741, 1450, 1382, 1224, 1170 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.41-6.20 (m, 4H), 6.18-6.13 (m, 1H), 6.06-5.99 (m, 1H), 5.85-5.71 (m, 2H), 5.18 (d, J = 17.2 Hz, 2H), 5.09 (d, J = 10.4 Hz, 2H) 4.31-4.22 (m, 2H), 3.72-3.62 (m, 4H), 3.67 (s, 6H), 1.48 (d, J = 7.3 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 173.39, 135.40, 133.99, 128.18 (dd, J_{CP} = 173.2, 5.0 Hz), 117.54, 53.67, 51.99, 47.01, 16.94; ³¹P NMR (162 MHz, CDCl₃) δ 16.69; LRMS calculated for C₁₈H₃₁N₂O₇P₂ (M+H)⁺ required 449.4, found 449.4.

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The characterization of the C_2 -symmetric alanine-derived vinyl phosphonamidic anhydride, single diastereomer (8a or 8b, bottom R_f) was as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.10 (EtOAc); FTIR 1741, 1450, 1382, 1224, 1170 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.41-6.20 (m, 4H), 6.18-6.13 (m, 1H), 6.06-5.99 (m, 1H), 5.85-5.71 (m, 2H), 5.15 (d, J = 16.9 Hz, 2H), 5.06 (d, J = 11.0 Hz, 2H), 4.60-4.49 (m, 2H), 3.72-3.62 (m, 4H), 3.66 (s, 6H), 1.45 (d, J = 7.3 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 173.39, 135.40, 134.11, 127.99 (d, J_{CP} = 178.8 Hz), 117.46, 52.83, 51.94, 46.35, 15.97; ³¹P NMR (162 MHz, CDCl₃) δ 16.70; LRMS calculated for $C_{18}H_{31}N_2O_7P_2$ (M+H)⁺ required 449.4, found 449.4.

VI. Preparation of Leucine-Derived Vinyl Phosphonamidic Anhydrides (9a-c)

A mixture of the diastereomeric vinylchloridates ($30P_SS$) and ($30P_RS$) (355 mg, 1.21 mmol) was subjected to the conditions described in Part II of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 135 mg (42%) of the pseudo-*meso* diastereomer (9c) and 126 mg (39%) of an inseparable mixture of C_2 -symmetric diastereomers (9a) and (9b), both as colorless oils.

The characterization of the pseudo-*meso* leucine-derived vinyl phosphonamidic anhydride (**9c**) was as follows: TLC R_f = 0.68 (EtOAc); $[\alpha]^{25}$ = -15.59 (c = 0.68, CHCl₃); FTIR 1740, 1649, 1461, 1438, 1387, 1207 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.45-6.22 (m, 4H), 6.19-6.11 (m, 1H), 6.06-5.98 (m, 1H), 5.82-5.72 (m, 2H), 5.13 (dd, J = 17.1, 1.2 Hz, 2H), 5.09 (dd, J = 17.1, 1.2 Hz, 2H), 4.46-4.40 (m, 1H), 4.33 (ddd, J = 13.2, 9.5, 5.3 Hz, 1H), 3.64 (s, 3H), 3.64-3.59 (m, 4H), 3.63 (s, 3H), 1.79-1.56 (m, 6H), 0.88 (d, J = 6.3 Hz, 6H), 0.87 (d, J = 6.2 Hz, 6H);

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¹³C NMR (100 MHz, CDCl₃) δ 173.44 (d, $J_{\rm CP}$ = 2.5 Hz), 173.14, 135.05, 135.05, 133.52, 133.52, 128.51 (dd, $J_{\rm CP}$ = 166.4, 8.2 Hz), 128.33 (dd, $J_{\rm CP}$ = 166.4, 9.0 Hz), 117.95, 117.68, 56.08 (d, $J_{\rm CP}$ = 3.4 Hz), 55.68 (d, $J_{\rm CP}$ = 4.3 Hz), 51.85, 51.76, 47.11 (d, $J_{\rm CP}$ = 5.2 Hz), 46.52 (d, $J_{\rm CP}$ = 5.2 Hz), 38.71 (d, $J_{\rm CP}$ = 3.6 Hz), 37.61 (d, $J_{\rm CP}$ = 3.0 Hz), 24.32, 24.06, 22.83, 22.83, 21.50, 21.34; ³¹P NMR (162 MHz, CDCl₃) δ 16.48 (d, $J_{\rm PP}$ = 37.5 Hz), 15.78 (d, $J_{\rm PP}$ = 37.5 Hz); HRMS calculated for C₂₄H₄₃N₂O₇P₂ (M+H)⁺ required 533.2546, found 533.2550.

Characterization of the C₂-symmetric leucine-derived vinyl phosphonamidic anhydrides (9a,b) as a mixture was as follows: TLC R_f = 0.24 (EtOAc); FTIR 1741, 1642, 1613, 1469, 1438, 1370, 1233 (P=O), 1207 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.37-6.21 (m, 8H), 6.08 (ddd, J = 10.2, 8.6, 4.2 Hz, 2H), 5.94 (ddd, J = 10.2, 8.5, 4.3 Hz, 2H), 5.75-5.65 (m, 4H), 5.11-4.96 (m, 8H), 4.40-4.29 (m, 4H), 3.65-3.51 (m, 8H), 3.57 (s, 6H), 3.56 (s, 6H), 1.74-1.49 (m, 12H), 0.87-0.81 (m, 24H); ¹³C NMR (100 MHz, CDCl₃) δ 173.05, 172.93, 135.08, 135.01, 133.92, 133.86, 128.18 (dd, $J_{\rm CP}$ = 176.8, 4.9 Hz), 127.98 (dd, $J_{\rm CP}$ = 174.9, 4.1 Hz), 117.59, 117.33, 55.67, 55.50, 51.66, 51.60, 46.62, 46.22, 38.32, 37.49, 24.17, 23.98, 22.71, 22.71, 21.13, 21.05; ³¹P NMR (162 MHz, CDCl₃) δ 17.46, 16.80; HRMS calculated for C₂₄H₄₃N₂O₇P₂(M+H)⁺ required 533.2546, found 533.2556.

VII. Preparation of Phenylalanine-Derived Vinyl Phosphonamidic Anhydrides (10a-c)

A mixture of the diastereomeric vinyl chloridates ($31P_SS$) and ($31P_RS$) (248 mg, 0.76 mmol) was subjected to the conditions described in Part I of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 61 mg (27%) of the pseudo-*meso* diastereomer (10c) and 64 mg (28%) of an inseparable mixture of C_2 -symmetric diastereomers (10a) and (10b), both as colorless oils.

Characterization of the pseudo-*meso* phenylalaninevinyl phosphonamidic anhydride (**10c**) was as follows: TLC R_f = 0.60 (EtOAc); $[\alpha]^{25}$ = -78.33 (c = 0.66, CHCl₃); FTIR 1740, 1439, 1379, 1242, 1166, 750, 681 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.31-7.12 (m, 10H), 6.34-6.15 (m, 2H), 6.12-5.94 (m, 1H), 5.82-5.55 (m, 5H), 5.17 (dd, J = 17.1, 1.0 Hz, 1H), 5.14 (dd, J = 17.1, 1.1 Hz, 1H), 5.11-5.06 (m, 2H), 4.69-4.59 (m, 2H), 3.64 (s, 3H), 3.64 (s, 3H), 3.61-3.57 (m, 2H), 3.52-3.48 (m, 2H), 3.36 (dd, J=12.1, 6.6 Hz, 1H), 3.32 (dd, J=12.0, 6.5 Hz, 1H), 3.09 (dd, J=14.5, 7.8 Hz, 1H), 3.07 (dd, J=14.6, 8.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 172.24, 172.12, 137.52, 137.49, 134.57, 134.45, 133.66, 133.55, 129.27, 129.25, 128.32 (dd, J_CP=181.2,

7.2 Hz), 128.30, 128.26, 127.17 (dd, $J_{\rm CP}$ = 184.4, 6.1 Hz), 126.44, 126.42, 118.30, 118.22, 58.71, 58.49, 51.89, 51.77, 47.26, 47.02, 35.85, 35.71; ³¹P NMR (162 MHz, CDCl₃) δ 15.90; HRMS calculated for $C_{30}H_{39}N_2O_7P_2$ (M+H)⁺ required 601.2233, found 601.2233.

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Characterization of the C_2 -symmetric phenylalanine-derived vinyl phosphonamidic anhydride, single diastereomer (**10a** or **10b**, top R_f) was as follows (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.30 (EtOAc); FTIR 1740, 1437, 1239, 1168, 750, 681 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.28-7.09 (m, 10H), 6.18-5.43 (m, 8H), 5.16-5.01 (m, 4H), 4.71-4.63 (m, 1H), 4.54-4.46 (m, 1H), 3.59 (s, 6H), 3.55-3.47 (m, 2H), 3.36-3.26 (m, 4H), 3.13-3.03 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 171.90, 137.31, 134.41, 133.63, 129.05, 128.05 (dd, J_{CP} = 177.5, 4.4 Hz), 127.99, 126.18, 117.87, 58.64, 51.66, 47.18, 35.40; ³¹P NMR (162 MHz, CDCl₃) δ 16.98; LRMS calculated for C₃₀H₃₉N₂O₇P₂ (M+H)⁺ required 601.3, found 601.3.

The characterization of the C_2 -symmetric phenylalanine-derived vinyl phosphonamidic anhydride, single diastereomer (**10a** or **10b**, bottom R_f) yielded the following (contamination with other diastereomer does not allow for optical rotation measurement): TLC $R_f = 0.30$ (EtOAc); FTIR 1740, 1437, 1239, 1168, 750, 681 cm⁻¹; 1 H NMR (400 MHz, CDCl₃) δ 7.28-7.09 (m, 10H), 6.18-5.43 (m, 8H), 5.16-5.01 (m, 4H), 4.71-4.63 (m, 1H), 4.54-4.46 (m, 1H), 3.58 (s, 6H), 3.55-3.47 (m, 2H), 3.36-3.26 (m, 4H), 3.13-3.03 (m, 2H); 13 C NMR (100 MHz, CDCl₃) δ 172.05, 137.18, 134.45, 133.75, 128.96, 128.12, 127.39 (dd, J_{CP} =177.0, 4.6 Hz), 126.27, 117.96, 58.14, 51.64, 46.84, 35.69; 31 P NMR (162 MHz, CDCl₃) δ 17.56; LRMS calculated for $C_{30}H_{39}N_2O_7P_2$ (M+H) $^+$ required 601.3, found 601.3.

VIII. Preparation of Valine-derived Vinyl Phosphonamidic Anhydrides (11a-c)

A mixture of the diastereomeric vinyl chloridates ($32P_SS$) and ($32P_RS$) (734 mg, 2.63 mmol) was subjected to the conditions described in Part I of this example. Flash chromatography (SiO₂, 1:1 Hex/EtOAc) afforded 241 mg (40%) of the pseudo-*meso* diastereomer (11c) and 266 mg (44%) of an inseparable mixture of C_2 -symmetric diastereomers (11a) and (11b), both as colorless oils.

The characterization of the pseudo-*meso* valine-derived vinyl phosphonamidic anhydride (11c) was as follows: TLC $R_f = 0.50$ (EtOAc); $[\alpha]^{25} = -73.42$ (c = 0.54, CHCl₃); FTIR 1738, 1479, 1371, 1247, 1203 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.38-6.20 (m, 4H), 6.17-6.09 (m,

1H), 6.03-5.96 (m, 1H), 5.79 (dddd, J = 16.9, 10.1, 6.8, 6.8 Hz, 2H), 5.12 (d, J = 17.1 Hz, 2H), 5.01 (dd, J = 10.0, 1.5 Hz, 2H), 3.92 (t, J = 11.3 Hz, 1H), 3.86 (t, J = 11.0 Hz, 1H), 3.81-3.70 (m, 2H), 3.70-3.58 (m, 2H), 3.64 (s, 3H), 3.63 (s, 3H), 2.77-2.17 (m, 2H), 0.94 (d, J = 6.7 Hz, 3H), 0.90 (d, J = 6.7 Hz, 3H), 0.88 (d, J = 6.4 Hz, 3H), 0.87 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.02 (d, J_{CP} = 2.9 Hz), 171.7 (d, J_{CP} = 1.3Hz), 135.14, 135.14, 134.07, 133.24, 128.75 (dd, J_{CP} = 182.6, 9.0 Hz), 128.12 (dd, J_{CP} = 183.9, J_{CP} = 8.9 Hz), 117.67, 117.57, 63.71 (d, J_{CP} = 4.1 Hz), 63.59 (d, J_{CP} = 3.0 Hz), 51.40, 51.31, 46.45 (d, J_{CP} = 4.8 Hz), 46.16 (d, J_{CP} = 4.0 Hz), 27.71 (d, J_{CP} = 3.3 Hz), 27.13 (d, J_{CP} = 2.6 Hz), 19.77, 19.65, 19.59, 19.46; ³¹P NMR (162 MHz, CDCl₃) δ 16.32 (d, J_{PP} = 35. Hz), 15.98 (d, J_{PP} = 35.9 Hz); HRMS calculated for C₂₂H₃₉N₂O₇P₂ (M+H)⁺ required 505.2223, found 505.2227.

The characterization of the C_2 -symmetric valine-derived vinyl phosphonamidic anhydride, single diastereomer (**11a** or **11b**, top R_f) yielded the following (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.40 (EtOAc); FTIR 1738, 1436, 1371, 1243, 1204 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.36-6.22 (m, 4H), 6.19-6.10 (m, 1H), 6.03-5.97 (m, 1H), 5.82-5.71 (m, 2H), 5.15-5.02 (m, 4H), 3.94-3.87 (t, J = 11.8 Hz, 1H), 3.83 (t, J = 11.7 Hz, 1H), 3.79-3.67 (m, 2H), 3.69-3.57 (m, 2H), 3.64 (s, 6H), 2.30-2.17 (m, 2H), 0.90 (d, J = 6.5 Hz, 6H), 0.87 (d, J = 6.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 171.90, 135.06, 134.61, 127.88 (dd, J_{CP} = 175.2, 4.6 Hz), 117.72, 63.34, 51.44, 46.25, 27.68, 19.67, 19.42; ³¹P NMR (162 MHz, CDCl₃) δ 16.87; LRMS calculated for $C_{22}H_{39}N_2O_7P_2$ (M+H)⁺ required 505.5, found 505.5.

Characterization of the C_2 -symmetric valine-derived vinyl phosphonamidic anhydride, single diastereomer (**11a** or **11b**, bottom R_f) resulted in the following (contamination with other diastereomer does not allow for optical rotation measurement): TLC R_f = 0.40 (EtOAc); FTIR 1738, 1436, 1371, 1243, 1204 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.36-6.22 (m, 4H), 6.19-6.10 (m, 1H), 6.03-5.97 (m, 1H), 5.82-5.71 (m, 2H), 5.15-5.02 (m, 4H), 3.94-3.87 (t, J= 11.8 Hz, 1H), 3.83 (t, J= 11.7 Hz, 1H), 3.79-3.67 (m, 2H), 3.69-3.57 (m, 2H), 3.62 (s, 6H), 2.30-2.17 (m, 2H), 0.95 (d, J= 6.6 Hz, 6H), 0.87 (d, J= 6.5 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 171.90, 135.10, 134.02, 128.40 (dd, J_{CP} = 174.5, 3.9 Hz), 117.56, 63.62, 51.39, 45.99, 27.09, 19.72, 19.53; ³¹P NMR (162 MHz, CDCl₃) δ 17.20; LRMS calculated for C₂₂H₃₉N₂O₇P₂ (M+H)⁺ required 505.5, found 505.5.

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EXAMPLE 3

Scheme F depicts the general overall reaction scheme followed in Parts I-II below, as well as the various compounds which can be prepared according to the procedure described in this example.

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Scheme F

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$$MeO_2C$$

$$\begin{array}{c} Bn \\ N \\ P \\ O \\ P \\ N \end{array}$$

$$\begin{array}{c} O \\ Bn \\ CO_2Me \\ \hline \\ MeO_2C \\ \hline \\ Bn \\ Bn \\ \end{array}$$

$$\begin{array}{c} O \\ O \\ P \\ N \\ CO_2Me \\ \hline \\ 10c \\ \end{array}$$

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I. Preparation of Pseudo-meso Bicyclic Leucine-derived Phosphonamidic Anhydride (13c) Leucine-derived vinyl phosphonamidic anhydride (9c) (88 mg, 0.165 mmol) and CH₂Cl₂ (15 mL) were added to a flame-dried 25 mL round bottom flask. The mixture was stirred, and the system was purged with argon for 10 minutes using a gas aerating tube. The Grubbs Catalyst 1 (6.8 mg, 8 μmol) was added under argon, and the reaction mixture was stirred and monitored for disappearance of the starting material. Upon completion, the reaction was concentrated under reduced pressure, passed through a plug of silica using EtOAc, and further concentrated under reduced pressure to give a crude oil. Flash chromatography (SiO₂, 100% EtOAc) afforded the bicyclic phosphonamidic anhydride (13c) (75 mg, 96%) (see Scheme G) as a colorless oil.

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Scheme G

MeO₂C P N CO₂Me

13c

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Characterization of the pseudo-meso bicyclic leucine-derived phosphonamidic anhydride (13c) was as follows: TLC $R_f = 0.20$ (EtOAc); $[\alpha]^{25} = +11.1$ (c = 0.19, CHCl₃); FTIR 1742, 1587, 1451, 1390, 1346, 1241 (P=O), 1199 (P=O) cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.11-7.06 (m, 1H), 7.00-6.95 (m, 1H), 6.25 (dd, J= 30.3, 9.0 Hz, 1H), 6.12 (dd, J= 30.2, 9.0 Hz, 1H), 4.47 (ddd, J_{HP} = 9.4 Hz, J_{HH} = 6.5, 6.5 Hz, 1H), 4.29 (ddd, J_{HP} = 7.2 Hz, J_{HH} = 7.2, 7.2 Hz, 1H), 4.17-4.06 (m, 2H), 3.76-3.67 (m, 2 H), 3.67 (s, 3H), 3.65 (s, 3H), 1.67-1.64 (m, 4H), 1.64-1.51 (m, 2H), 0.97 (d, J= 6.3 Hz, 3H), 0.93-0.90 (m, 9H); 13 C NMR (100 MHz, CDCl₃) δ 173.60 (d, J_{CP} = 1.2 Hz), 173.13 (d, J_{CP} = 1.9 Hz), 146.15 (d, J_{CP} = 17.0 Hz), 145.75 (d, J_{CP} = 17.4 Hz), 119.24 (d, J_{CP} = 161.9 Hz), 118.62 (dd, J_{CP} = 166.2, 2.9 Hz), 52.20 (d, J_{CP} = 4.1 Hz), 52.04 (d, J_{CP} = 4.5 Hz), 51.84, 51.73, 47.18 (d, J_{CP} = 31.8 Hz), 46.82 (d, J_{CP} = 31.1 Hz), 39.14 (d, J_{CP} = 3.4 Hz), 38.49 (d, J_{CP} = 3.8 Hz), 24.48, 24.47, 23.06, 22.99, 21.28, 21.07; 31 P NMR (162 MHz, CDCl₃) δ 33.55 (d, J_{PP} = 24.0 Hz), 32.38 (d, J_{PP} = 24.0 Hz); HRMS calculated for $C_{20}H_{35}N_2O_7P_2$ (M+H)⁺ required 477.1919, found 477.1911.

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II. Preparation of the Pseudo-meso Bicyclic Phenylalanine-Derived Phosphonamidic Anhydride (14c)

In a procedure similar to the preparation of the bicyclic leucine-derived phosphonamidic anhydrides (13c) as described in Part I of this example, compound (10c) (102 mg, 0.170 mmol) was subjected to Grubbs Catalyst 1 (7.0 mg, 8 μ mol) in 15 mL CH₂Cl₂. Flash chromatography (SiO₂, 100% EtOAc) afforded (14c) (86 mg, 93%) (see Scheme H) as a colorless oil.

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Scheme H

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14c

Characterization of the resulting pseudo-meso bicyclic phenylalanine-derived phosphonamidic anhydride (14c) yielded the following: TLC R_f = 0.37 (EtOAc); [α]²⁵ = -5.4 (c = 0.74, CHCl₃); FTIR 2949, 2923, 2854, 1740, 1454, 1439, 1348, 1318, 1239, 1207, 1177, 901, 749, 698 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.32-7.18 (m, 10H), 7.08-7.03 (m, 1H), 6.96-6.91 (m, 1H), 6.21 (dd, J_{HP} = 30.5 Hz, J_{HH} = 9.0 Hz, 1H), 5.94 (dd, J_{HP} = 30.2 Hz, J_{HH} = 8.9 Hz, 1H), 4.66 (ddd, J_{HP} = 7.7 Hz, J_{HH} = 7.7, 7.7 Hz, 1H), 4.59 (ddd, J_{HP} = 7.8 Hz, J_{HH} = 7.8, 7.8 Hz, 1H), 4.16-4.09 (m, 2H), 3.88-3.77 (m, 2H), 3.63 (s, 3H), 3.86 (s, 3H), 3.32 (dd, J = 14.1, 7.8, Hz, 1H), 3.27 (dd, J = 13.3, 7.1 Hz, 1H), 3.09 (dd, J = 13.7, 8.0 Hz, 1H), 3.03 (dd, J = 14.0, 7.7 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 172.14, 171.95, 146.12 (d, J_{CP} = 11.5 Hz), 145.95 (d, J_{CP} = 11.7 Hz), 136.82, 136.26, 129.19, 128.79, 128.53, 128.46, 126.82, 126.77, 119.19 (d, J_{CP} = 164.4 Hz), 118.7 (d, J_{CP} = 163.7 Hz), 56.04 (d, J_{CP} = 4.8 Hz), 55.00 (d, J_{CP} = 4.3 Hz), 52.01, 51.91, 48.29 (d, J_{CP} = 31.3 Hz), 47.62 (d, J_{CP} = 30.9 Hz), 37.05, 36.58 (d, J_{CP} = 3.7 Hz); ³¹P NMR (162 MHz, CDCl₃) δ 33.21 (d, J_{PP} = 24.9 Hz), 32.24 (d, J_{PP} = 24.9 Hz); HRMS calculated for $C_{26}H_{31}N_{2}O_{7}P_{2}$ (M+H)⁺ required 545.1606, found 545.1589.

In Parts I-II of this procedure, methylene chloride was the solvent utilized. However, toluene, benzene, chlorobenzene, dichlorobenzene, DME, and mixtures thereof are also suitable solvents. Furthermore, while Grubbs Catalyst 1 was used as the catalyst, it will be appreciated that Grubbs Catalyst 2 or 3 could also be utilized. Finally, while the procedure was carried out at a temperature of 40°C, temperatures of from about 15-80°C would also be suitable.

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We Claim:

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1. A compound according to a formula selected from the group consisting of

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and

$$\begin{array}{c|c}
 & O & O \\
 & // & // \\
 & P & O & P \\
 & P & N \\
 & R^2O_2C & R^1 & R^1 & CO_2R^2
\end{array}$$

wherein:

each X is individually selected from the group consisting of oxygen, -NH, and -NOR¹;

each R¹ is individually selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

each R² is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups, 2-15 mer peptides, and benzyl groups.

2. The compound of claim 1, wherein at least one R¹ comprises an amino acid side chain selected from the group consisting of

wherein each R³ is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups, and benzyl groups.

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- 3. The compound of claim 1, wherein:
 - each R^1 is individually selected from the group consisting of -CH₃, $CH_2CH(R^4)_2$, -CH₂R⁴, and -CH(R⁴)₂, with each R⁴ being individually selected from the group consisting of alkyl groups, aryl groups, and benzyl groups; and

each R² is individually selected from the group consisting of -CH₃ and -CHCH₂.

4. The compound of claim 3, wherein each R⁴ is individually selected from the group consisting of methyl groups and phenyl groups.

5. The compound of claim 1, wherein said compound comprises a formula selected from the group consisting of

$$MeO_2C$$
 Me_2HC $CHMe_2$ and MeO_2C Ph Ph CO_2Me

6. A method of forming a phosphonamide compound comprising the steps of: reacting an allylated compound with a phosphonic compound in order to form an intermediate compound, said allylated compound being according to the formula

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wherein:

R1 is selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

R² is selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups 2-15 mer peptides, and benzyl groups,

and said phosphonic compound being according to the formula R⁴POY₂,

wherein:

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R⁴ is selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups 2-15 mer peptides, and benzyl groups; and

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each Y is individually selected from the group consisting of the halogens; and

dimerizing said intermediate compound to yield the phosphonamide compound.

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7. The method of claim 6, wherein said allylated compound is according to the formula

$$R^1$$
 OR^2

wherein R¹ is selected from the group consisting of -CH₃, -CH₂CHMe₂, -CH₂Ph, and -10 CHMe₂, and R² is an alkyl group.

8. The method of claim 6, wherein said phosphonic compound is according to the formula R⁴POCl₂, wherein R⁴ is selected from the group consisting of -CH₃ and -CHCH₂.

9. The method of claim 6, wherein said intermediate compound is according to the formula

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$$R^2$$
 P N R^1 OR^2

and

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wherein:

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R¹ is selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides;

R² is selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups 2-15 mer peptides, and benzyl groups; and

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each Y is individually selected from the group consisting of the halogens.

10. The method of claim 6, wherein said phosphonamide compound is according to the formula

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wherein:

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- each X is individually selected from the group consisting of oxygen, -NH, and -NOR1;
- each R¹ is individually selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

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each R² is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups, 2-15 mer peptides, and benzyl groups.

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11. The method of claim 6, wherein said reacting step is carried out in the presence of a compound selected from the group consisting of bases, catalysts, and solvents.

12. The method of claim 11, wherein said compound is a base selected from the group consisting of Et₃N, pyridine, NaHCO₃, Na₂CO₃, K₂CO₃, NaH, KH, and mixtures thereof.

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13. The method of claim 11, wherein said compound is a catalyst and said catalyst is DMAP.

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14. The method of claim 11, wherein said compound is a solvent selected from the group consisting of acetonitrile, chloroform, toluene, benzene, THF, diethyl ether, DME, and mixtures thereof.

- 15. The method of claim 6, wherein said reacting step and said dimerizing step are each individually carried out at a temperature of from about -20-20°C.
- 16. The method of claim 6, wherein said reacting step is carried out in the presence of a base.
 - 17. The method of claim 16, wherein said base is selected from the group consisting of Et₃N, pyridine, NaHCO₃, Na₂CO₃, K₂CO₃, NaH, KH, and mixtures thereof.
- 18. The method of claim 6, further including the step of reacting said phosphonamide compound with a ring-closing catalyst to yield a bicyclic phosphonamide.
 - 19. The method of claim 18, wherein said ring-closing catalyst is a Grubbs catalyst.
- 15 20. The method of claim 19, wherein said Grubbs catalyst is selected from the group consisting of

21. The method of claim 18, wherein said bicyclic phosphonamide is represented by the formula

$$R^2O_2C$$
 R^1
 R^1
 CO_2R^2

wherein:

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each R¹ is individually selected from the group consisting of hydrogen, substituted and unsubstituted amino acid side chains, and 2-15 mer peptides; and

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each R² is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups, 2-15 mer peptides, and benzyl groups.

22. The method of claim 21, wherein at least one R¹ comprises an amino acid side chain selected from the group consisting of

wherein each R³ is individually selected from the group consisting of hydrogen, branched and unbranched alkyl groups, branched and unbranched alkenyl groups, branched and unbranched alkynyl groups, allyl groups, aryl groups, acyl groups, and benzyl groups.

INTERNATIONAL SEARCH REPORT

Intermonal application No.
PCT/US01/41602

A. CLASSIFICATION OF SUBJECT MATTER IPC(7) : CO7F 9/553, 9/572, 9/22, 9/06, 9/02				
US CL: 562/878; 560, 19 According to International Patent Classification (IPC) or to both national classification and IPC				
B. FIELDS SEARCHED				
	ocumentation searched (classification system followed	by classification symbols)		
U.S. :	562/878; 560, 19	,	į	
Documentat	ion searched other than minimum documentation to the	extent that such documents are included in	n the fields searched	
Please See Extra Sheet.				
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)				
C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages	Relevant to claim No.	
A	US 2,502,966 A (KOSOLAPOFF) document.	04 April 1950, see entire	1-5	
A	US 3,248,458 A (OERTEL et al.) document.	26 April 1966, see entire	1-5	
A	US 3,366,714 A (POPOFF) 30 January	y 1968, see entire document.	1-5	
A	US 3,972,866 A (FORTUIN et al.) 03 August 1976, see entire document.		1-5	
			·	
Furth	ner documents are listed in the continuation of Box C.	See patent family annex.		
* Sp	ecial categories of cited documents:	"T" later document published after the integrated date and not in conflict with the applications."	ernational filing date or priority	
	cument defining the general state of the art which is not considered be of particular relevance	the principle or theory underlying the		
	rlier document published on or after the international filing date	"X" document of particular relevance; the considered novel or cannot be considered.	e claimed invention cannot be red to involve an inventive step	
	cument which may throw doubts on priority claim(s) or which is ed to establish the publication date of another citation or other	when the document is taken alone	/	
	ecial reason (as specified)	"Y" document of particular relevance; the considered to involve an inventive	step when the document is	
me	cument referring to an oral disclosure, use, exhibition or other	combined with one or more other such being obvious to a person skilled in	the art	
	cument published prior to the international filing date but later than priority date claimed	"&" document member of the same paten		
Date of the actual completion of the international search Date of mailing of the international search report		arch report		
10 OCTO	DBER 2001	250CT2001		
Commissioner of Patents and Trademarks		Authorized officer	MAD -	
Box PCT Washington, D.C. 20231		FLOYD D. HIGEL		
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INTERNATIONAL SEARCH REPORT

Inter_____il application No. PCT/US01/41602

B. FIELDS SEARCHED Documentation other than minimum documentation that are included in the fields searched:			
CHEMICAL ABSTRACTS INDEX CHEMICAS CURRENT ABSTRACT OF CHEMISTRY			
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